DNA PRECIPITATION

Preparation:

1. Lightly bleach work space and put on latex gloves.
2. Get samples (plate or tube format).
3. Get reagents from chemical room. Chill 100% ethanol in freezer.
4. Write in lab book date/time and what specimens you are working on.

Prepare Reagents:

1. Maintain 100% ethanol on ice.
2. Prepare 70% ethanol with ultra-clean H2O.
3. Check pH of 3M Na-Acetate (should be pH 5.2).

Precipitate DNA:

1. Add 1/10 volume of 3 M Na-Acetate pH 5.2, and 2 to 2.5 volumes of ice-cold 100% ethanol to the DNA sample
2. Mix, and store at -20°C for at least 1 hour to precipitate the DNA
3. Recover the precipitated DNA by centrifugation at full speed in a 4°C microcentrifuge for 15-20 minutes (in plate format, spin for 30 min at max speed at 4°C)

DNA clean-up:

1. Pour off the ethanol (tap onto paper towel if needed) and wash the pellet twice with room-temperature 70% ethanol (in 4°C microcentrifuge spin at 6000 x *g*, for plate spin at 1650 RCF 4°C for 15 min)
2. Pour off the ethanol (tap onto paper towel if needed) and allow the DNA pellet to air-dry

Elute:

1. Resuspend the DNA in a suitable volume of sterile TE buffer or distilled water